biosera

# Technical Data Sheet

12/07/2017

## Kanamycin solution 100X

Product code: SM-K0271

**Theoretical pH** : 8 ± 1

**Storage conditions** : -20°C

**Shelf life** : 24 month

**Sterility tests** :

- Bacteria in aerobic and anaerobic conditions
- Fungi and yeasts

**Endotoxin** : <10 EU/ml

**Composition** : 10 mg/ml Kanamycin A

9 mg/ml Sodium Chloride

**Recommended use** :

Use in cell culture applications at 10 ml/l. This concentration is for tissue culture media containing serum; serum-free media generally require lower concentration.

- Respect storage conditions of the product
- Do not use the product after its expiry date
- Store product in an area protected from light
- Manipulate the product in aseptic conditions (e.g. : under laminar air flow)
- Wear clothes adapted to the manipulation of the product to avoid contamination (e.g. : gloves, mask, hygiene cap, overall…)
- In order to preserve all product qualities, it is recommended to thaw out the flask, to aliquote, then to re-freeze the produced flasks rather than to thaw out and re-freeze the flask at each use.
- It is recommended to use the product immediately after its thaw out.

The product is intended to be used in vitro, in laboratory only. Do not use it in therapy, human or veterinary applications.

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**Stability:** 5 days at 37°C

## Mode of Action:

Binds to 70S ribosomal subunit; inhibits translocation; elicits miscoding.

## Antimicrobial spectrum:

Gram-negative and Gram-positive bacteria, and mycoplasma.

## Application:

Antibiotics, combined with good sterile technique, help prevent microbiological contamination
When an irreplaceable culture becomes contaminated, determine if the contamination is bacteria, fungus, mycoplasma, or yeast. Isolate the contaminated culture from other cell lines. Clean incubators and laminar flow hoods with a laboratory disinfectant, and check HEPA filters.
Kanamycin at high concentration can be toxic to some cell lines; therefore, perform a dose response test to determine the level at which Kanamycin becomes toxic.
The following is a suggested procedure for determining toxicity levels and decontaminating cultures.

1) Dissociate, count, and dilute the cells in antibiotic free medium. Dilute the cells to the concentration used for regular cell passage.
2) Dispense the cell suspension into a multiwell culture plate or several small flasks. Add the Kanamycin to each well in a range of concentrations.
3) Observe the cells daily for signs of toxicity such as sloughing, appearance of vacuoles, decrease in confluency, and rounding.
4) When the toxic level has been determinated, culture the cells for two to three passages using the Kanamycin at a concentration one to two fold lower than the toxic concentration.
5) Culture the cells for one passage in an antibiotic-free media
6) Repeat step 4.
7) Culture the cells in antibiotic-free medium for 4 to 6 passages to determine if the contamination has been eliminated.

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